

Cryptococcus

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1 ***Cryptococcus*: from environmental saprophyte to global**
2 **pathogen**

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Abstract

Cryptococcosis is a globally distributed invasive fungal infection caused by species within the genus *Cryptococcus* that presents substantial therapeutic challenges. Although natural human-to-human transmission has never been observed, recent work has unveiled multiple virulence mechanisms that allow cryptococci to infect, disseminate within and ultimately kill their human host. In this Review, we describe these recent discoveries that illustrate the intricacy of host-pathogen interactions and reveal new details about host immune responses that either help protect against disease or increase host susceptibility. In addition, we discuss how this improved understanding of both the host and the pathogen informs potential new avenues for therapeutic development.

Cryptococcosis has been recognized since 1894, when the pathologist Otto Busse and physician Abraham Buschke jointly identified *Cryptococcus* as the cause of a chronic granuloma of the tibial bone in a 31-year-old woman. However, human cryptococcosis only became recognized as a major health threat with the onset of the AIDS pandemic in the 1980s, in which these fungal infections became a common AIDS-defining illness in patients with catastrophically reduced T-cell function (**Box 1**). Although cryptococcosis is predominantly a disease of immunocompromised patients, a recent outbreak of cryptococcosis in otherwise healthy individuals in North America and Canada (now known as the Pacific Northwest Outbreak) has focused attention on the capacity of some lineages of the pathogen to act as primary pathogens (see below).

Since its identification, cryptococcosis has been attributed to a single fungal species, *Cryptococcus neoformans*. However, improved molecular methods led to a previous variety, *Cryptococcus neoformans* var. *gattii*, being classified as a novel species, *Cryptococcus gattii*, in 2002¹. More recently, whole-genome sequencing-based analyses have highlighted the complex evolutionary history of this group (**Box 2**) and led to a proposal to further split *C. neoformans* into two species (*C. neoformans* and *Cryptococcus deneoformans*) and *C. gattii* into a total of five species (*C. gattii*, *Cryptococcus bacillisporus*, *Cryptococcus deuterogattii*, *Cryptococcus tetragattii* and *Cryptococcus decagattii*)². However, as detailed biological comparisons between these five species have not been yet undertaken,

we have adopted the simpler distinction into the two species *C. gattii* and *C. neoformans* throughout this article.

***Cryptococcus* transmission and disease onset**

In the environment, cryptococci reside in diverse ecological niches (**Box 3**). Both *C. neoformans* and *C. gattii* are abundant in decaying material within hollows of various tree species, although *C. gattii* has been suggested to favour trees with waxier cuticles (such as *Pseudotsuga menziesii*)^{3, 4}. Furthermore, *C. neoformans* is globally distributed, whereas *C. gattii* has classically been viewed as a tropical or subtropical fungus. However, increased surveillance has now identified environmental reservoirs for *C. gattii* in the Northern USA, Canada and Northern Europe, indicating that this species may also have a wider ecological range than previously recognized.

C. neoformans is particularly abundant in avian excreta^{4,5} and its association with feral pigeons could be a major source of infection in densely populated urban areas. In addition, both *C. neoformans* and *C. gattii* are able to survive and replicate within free-living amoebae and soil nematodes and it is possible that these alternative hosts may have an important role in determining the distribution and virulence of different cryptococcal lineages around the world (**Box 3**).

With the exception of very rare iatrogenic⁶ or zoonotic⁷ transmission events, naturally acquired cases of cryptococcosis are believed to start with inhalation of fungal cells from the environment. Within the lung, *Cryptococcus* species can cause pneumonia in immunosuppressed patients, but in immunocompetent hosts the fungal cells are either cleared by the immune system or establish an asymptomatic latent infection. Upon subsequent immunosuppression, this latent infection can then disseminate to other tissues, most notably the central nervous system (CNS). Once established within the CNS, cryptococcosis causes an overwhelming infection of the meninges and brain tissue that is frequently accompanied by raised intracranial pressure; without rapid and effective treatment, CNS infection is invariably fatal. Despite intensive investigations, it remains unclear whether reactivation and dissemination of long-term latent pulmonary infection is a more important cause of

cryptococcosis in patients than *de novo* acquisition from the environment, but experiments in animal models indicate that both routes are capable of causing lethal disease.

Exposure to *C. neoformans* is common in humans, as most individuals produce antibodies against this fungal species by school age⁸. During active growth, cryptococcal cells are too large to penetrate deep into the human lung and thus the initial inoculum is believed to comprise either desiccated cells or spores. The relative contribution of these two cell types to the burden of disease remains unclear, largely due to technical challenges associated with generating and purifying spores. However, recent studies have demonstrated that lethal brain infections can develop from spore inocula, that spores are readily phagocytosed by host immune cells and, interestingly, that rising humidity dramatically increases spore viability^{9,10,11}. Thus, as with other fungal pathogens such as *Coccidioides immitis*, environmental conditions may be an important factor in regulating human cryptococcal exposure.

Cryptococcal pathogenesis

Traditional virulence factors produced by *Cryptococcus* (such as the capsule and melanin production) and changes in fungal growth due to the host temperature (37°C) have been previously reviewed in great detail (see for example references^{12,13}). Therefore, in this section of the Review, we will focus on recently emerging concepts in cryptococcal pathogenesis.

Fungal morphology. Whether derived from spores or yeast cells, upon inhalation into a mammalian host, all cryptococci transition to or maintain a yeast form. When grown under laboratory conditions, *Cryptococcus* cells are round and 5-7 µm in diameter. However, their cell size, structure, and characteristics can vary dramatically within the host.

The best-characterized atypical morphology of *Cryptococcus* cells is the titan cell¹⁴ (**Figure 1**). Titan cells are greater than 12 µm in diameter (excluding the capsule), polyploid, have highly cross-linked capsules and a thickened cell wall^{15,16}. Recent studies have shown that titan cells contain elevated levels of chitin. This polysaccharide is recognized and cleaved by host chitinases, which

induces a detrimental adaptive immune response (see below)¹⁷. Intriguingly, the polyploidy observed in titan cells enhances genetic adaptation to the stressful host environment, resulting in increased within-host survival¹⁸.

In addition to the large titan cells, unusually small cryptococcal cells have also been observed^{19,20} (**Figure 1**). These so-called “drop” or “micro” cells are only 2-4 µm in size, despite having a thickened cell wall, and appear adapted for growth within macrophages. At present, little is known about this cell type, although they appear to be relatively metabolically inactive and therefore may have an important role during the latent stage of disease.

In the environment or under laboratory conditions, cryptococci can also grow as hyphae (during sexual reproduction) or pseudohyphae, but (unlike other pathogenic fungi) these morphologies are not seen in human infections²¹. Recent studies overexpressing the transcription factor Znf2, a “master regulator” that triggers the transition from yeast to hyphal growth, showed that the hyphal form elicits a robust protective immune response and is readily cleared by the host^{22,23}, perhaps explaining why filamentous morphologies are not seen in mammalian infections. Interestingly, however, hyphal cryptococci are protected from predation by free-living amoebae²⁴ and thus mammalian and amoebal hosts presumably exert opposing selective pressures on this aspect of cryptococcal morphology (with mammalian hosts favouring the existence of the yeast forms and amoebae favouring hyphal forms).

Fungal ageing. Even within a clonal infection, not all cryptococcal cells are equal. For example, the age of individual cryptococcal cells has emerged as a factor that impacts survival in the host and subsequent pathogenesis²⁵. Older cells present in the initial infection, referred to as founder cells, are better able to resist phagocytosis and killing by phagocytes and are resistant to antifungal drugs. This increased resistance to phagocyte killing and antifungals is potentially due to changes in cell wall structure²⁶, and results in the accumulation of founder cells in the brain at a higher frequency than young cells²⁷.

Population-wide signals. In bacterial infections, quorum sensing is a well-known mechanism that regulates virulence according to population density. Interestingly, emerging data suggest that quorum sensing may also have an important role during cryptococcal pathogenesis. For example, a quorum sensing effect, mediated by an oligopeptide with 11 amino acids, was identified using mutations in the global repressor TUP1. Notably, although TUP1 is present in several species, the quorum sensing effect mediated by this oligopeptide appears only to occur in *C. neoformans*²⁸. However, more recently a different signaling molecule, pantothenic acid, has been demonstrated to mediate quorum sensing both between different cryptococcal strains and between cryptococci and other, relatively distantly related, fungal species²⁹. The adhesin Cfl1 has also been shown to modulate colony morphology in a paracrine manner³⁰. Activation of the hyphal regulator Znf2 (discussed above) induces expression of this adhesin, some of which is shed into the environment and triggers neighboring cells to activate Znf2, leading to a positive feedback loop. Thus cryptococci may communicate locally using a range of chemical messengers³¹.

Perhaps most unique is the observation that light-sensing pathways may also be important for virulence in *Cryptococcus* since deletion of either *Bwc1* or *Bwc2*, which encode two transcription factors that control fungal responses to light, reduces virulence in a murine model of infection³². In the dark, BWC1 and BWC2 bind to DNA and repress genes involved in filamentation. However, upon light activation, they release this inhibition leading to filamentation and upregulation of UV-resistance pathways. Thus, it is possible that an additional function of these two proteins is to detect darkness and prevent inappropriate filamentation within the host, which would induce a potent immune response and pathogen clearance.

Host immunity and pathogen subversion

One of the most remarkable discoveries of recent years has been the extent to which cryptococci are able to manipulate the host immune response to dampen inflammation, avoid killing by phagocytic cells and ultimately disseminate into the CNS.

Inflammatory perturbation. In general, environmental fungi trigger a potent inflammatory response upon entry into the human host. By contrast, cryptococci appear to be immunologically inert, driving much lower levels of inflammatory cytokine release *in vitro* than other human fungal pathogens such as *C. albicans*³³. This immunological masking relies on a variety of pathogen traits (**Figure 1**).

Firstly, the complex carbohydrates glucuronoxylomannan (GXM) and galactoxylomannan (GalXM), which make up most of the cryptococcal capsule, are extensively shed during infection and directly dampen inflammation by suppressing the pro-inflammatory NF- κ B pathway and driving down levels of pro-inflammatory cytokines such as TNF³⁴. In addition, emerging data indicate that cryptococcal chitin, and derivatives thereof, can also act to alter host inflammatory responses during infection¹⁷. Secondly, *Cryptococcus* blocks dendritic cell maturation by reducing both MHC class II-dependent antigen presentation and inhibiting the production of the pro-inflammatory cytokines interleukin (IL)-12 and IL-23³⁵. Lastly, via a series of as-yet poorly characterized steps, cryptococci are able to partially “repolarize” the immune response, at least in mice, from a strong Th1 response towards a weaker Th1 or often a Th2 response that is less effective at fungal clearance^{17,36-38}.

Collectively, these mechanisms generate an environment that is dominated by anti-inflammatory markers such as IL-4 and IL-33^{39,40,41} which, as a consequence, reduce cryptococcal killing by the immune system^{38,42}. Therefore, modulating natural immune responses to cryptococcal infection towards a more pro-inflammatory profile offers one potential avenue for treatment. However, such approaches need to be carefully managed in order to avoid the potentially fatal “immune overreactions” that can accompany overt inflammation, which can be just as life-threatening as the original infection (**Box 4**).

Avoidance and escape from phagocytes. Following entry into the lung, the first immune cell typically encountered by cryptococci is a phagocyte such as an alveolar macrophage or dendritic cell. However, cryptococci are predisposed to avoid killing by these cells, due to their long evolutionary history of exposure to environmental amoebae (**Box 3**). Several cryptococcal virulence factors such as capsule synthesis, melanization and urease secretion combine to protect the

fungus from the harsh environment within phagocytic cells by neutralizing reactive oxygen species and pH, allowing it to survive and proliferate within such cells (**Figure 2**)⁴³.

More recently, it has also become clear that cryptococci exhibit a remarkable strategy to escape from within phagocytes. This process, which has been labeled vomocytosis or extrusion, involves inducing the fusion of the phagosomal membrane with the plasma membrane, which results in the expulsion of the fungi from the phagocyte⁴⁴⁻⁴⁸. In addition, either this process, or a closely related one, can drive the direct “lateral transfer” of cryptococci between host cells^{44,45}. However, the underlying mechanisms of both of these remarkable processes remain unknown.

Although cryptococci employ several mechanisms to resist phagocytosis (such as through production of titan cells^{15,49} and the assembly of a thick polysaccharide capsule), fungal uptake by phagocytes can still occur. However, if uptake does occur, cryptococci perturb both phagosome maturation⁵⁰ and modify the phagosome membrane in order to allow nutrient exchange and ultimately escape from within the host cell^{51,52}. Notably, these effects are dependent on fungal virulence factors such as laccase and phospholipase B1. These enzymes have been classically thought of as having direct structural roles in melanin synthesis and membrane lipid modification, respectively, but the observation that they also mediate escape from phagocytosis suggests that laccase and phospholipase B1 may also have more subtle roles in modifying host signaling events^{36,53,54}.

Dissemination and entry into the CNS. A key feature of cryptococcal pathogenesis involves the exit of *Cryptococcus* from the lungs into peripheral blood circulation and entry into the CNS compartment. The CNS is both an immune privileged site and a highly sterile environment and thus *Cryptococcus* must have evolved potent methods to traverse the blood-brain barrier (BBB) and subsist in the CNS.

There are three proposed mechanisms that *Cryptococcus* could utilize to penetrate this impervious barrier. First, the yeasts could force their way between the tight junctions of the endothelial cells in a process known as

paracytosis, by using proteases such as Mpr1 to promote transmigration⁵⁵ (**Figure 2**). Impressively, when the *MPR1* gene was introduced into *Saccharomyces cerevisiae*, a fungus not normally able to penetrate the BBB, *S. cerevisiae* gained the ability to cross endothelial cells in an *in vitro* transwell assay, although the target of Mpr1 remains unknown. Additional studies utilizing powerful intravital imaging techniques demonstrated that cryptococci cross the BBB by inducing an embolic event in the microvasculature that lines the brain⁵⁶. In essence, the initial “capture” of yeast within the brain is therefore passive, with the relatively large yeast cells becoming trapped at points where the blood vessel narrows. However, following the initial passive arrest, cryptococcal migration into the brain tissue is an active process, since it occurs only with live fungal cells and is dependent on the secretion of the cryptococcal enzyme urease⁵⁷. To date, the part played by urease in this process remains enigmatic, although since urease produces ammonia, which is toxic towards mammalian cells, it is possible that urease acts to locally weaken the endothelial vessel wall, facilitating fungal entry.

The second mechanism of BBB penetration is transcytosis⁵⁸ (**Figure 2**). Hyaluronic acid situated on the surface of the cryptococcal cell binds to CD44 on the luminal endothelium, attaching the fungus to the host cell⁵⁹. This binding then induces protein kinase C-dependent actin remodeling in the host cell, leading it to engulf the attached *Cryptococcus*⁶⁰. Interestingly, recent work has revealed that the high levels of inositol present in the brain act as a trigger for this process, increasing hyaluronic acid expression in the fungus⁶¹.

Finally, *Cryptococcus* is postulated to cross the BBB by a third method involving “hitchhiking” within host phagocytes, in a process termed the “Trojan Horse” hypothesis (**Figure 2**). This hypothesis is supported by the observation that depletion of alveolar macrophages in mice significantly reduces cryptococcal dissemination to the CNS⁶², while infecting monocytes *in vitro* and transferring the cells into naïve hosts substantially increases cryptococcal accumulation in the brain compared to transferring *Cryptococcus* directly; both studies support the notion that phagocytes act as fungal carriers that breach the BBB⁶³. Although paracytosis, transcytosis, and Trojan Horse models are all fundamentally different, it is reasonable to conclude that elements of each of

these models are readily observed and likely occur in concert during natural infection.

Not much is known about the physiology of *Cryptococcus* after it has traversed the BBB. However, a recent study of the transcriptome of cryptococcal yeasts isolated from cerebrospinal fluid (CSF) samples of patients offers some clues⁶⁴. Most notably, *Cryptococcus* is remarkably metabolically active in the CSF *in vivo*, showing strong up-regulation of stress response genes and genes encoding enzymes that are involved in core metabolic processes; this is somewhat surprising, given that the CSF is a relatively nutrient depleted medium. By contrast, *Cryptococcus* growing in *ex vivo* CSF does not seem to be metabolically active, suggesting that the permanent cycling of CSF *in vivo* leads to a significantly higher nutrient content in the CSF than suggested by the analysis of *ex vivo* samples.

The modified fungal metabolism observed within the CSF is likely to have significant implications for pathogenesis. For instance, capsule synthesis is energetically highly demanding and there is a positive correlation between capsular size and severity of clinical disease⁶⁵. Therefore, these data suggest that yeasts in a more active metabolic state may drive more aggressive CNS infections. Furthermore, fungal cells in different metabolic states are likely to give rise to different immune responses, which may also impact disease severity. In agreement with this possibility, the presence of a CSF inflammatory response consisting of an interplay of robust Th1 (IFN- γ and IL-6), Th2 (IL-4 and IL-10) and Th17 (IL-17) cytokines has recently been shown to be highly predictive of more rapid clearance of infection and consequently improved survival in patients with HIV-associated cryptococcal meningitis⁶⁶ (**Box 4**).

Division of labour. The extent to which cryptococci can exploit phagocytic cells as a host has been strongly highlighted by the unusual cluster of cryptococcal disease now known as the Pacific Northwest Outbreak⁶⁷. Although cryptococcosis is typically a disease of immunocompromised hosts, almost all of the human and animal cases within the Pacific Northwest Outbreak were immunocompetent hosts who became infected with near clonal strains of *C. gattii* from the VGII lineage. Both the epidemiology and etiology of these

infections differ from “classical” cryptococcosis (typically caused by *C. neoformans* in HIV-positive individuals)^{33,68}, which has led to vigorous efforts in order to establish the underlying mechanism driving virulence in the *C. gattii* VGII lineage.

The ability of the *C. gattii* VGII lineage to establish disease in individuals with a fully functional immune system seems to stem from a capacity to replicate extremely rapidly within host phagocytes (**Figure 2**), presumably overwhelming the host before adaptive immunity can be triggered⁶⁹. Recent data has revealed that this rapid proliferation is, in turn, driven by a remarkable “division of labour” mechanism. In response to reactive oxygen species generated by the phagocyte, intracellular cryptococcal cells adopt different fates; some cryptococcal cells cease growth and acquire an unusual morphology characterized by extensive tubularisation of their mitochondria, whereas neighboring cells do not undergo this morphological transition. Notably, via a mechanism that remains unclear, the cells that undergo the morphological switch then protect neighboring cryptococci from the antimicrobial activity of the host phagocyte, enabling these cells to replicate rapidly, maximizing the proliferative capacity of the population as a whole⁷⁰. These data highlight the *Cryptococcus*—phagocyte interaction as a key aspect of infection that may offer powerful opportunities for therapeutic intervention in both *C. neoformans* and *C. gattii* infections.

Anti-cryptococcal therapeutics

Despite its global distribution, treatment of cryptococcosis remains a major challenge, relying on a limited arsenal of decades-old therapeutic agents. Furthermore, therapeutic outcomes are generally poor and even with amphotericin-based therapy (to target *Cryptococcus*) and widespread access to anti-retroviral therapy (to target HIV, since most patients are immunocompromised HIV-positive patients), acute (3-month) mortality following cryptococcal meningoencephalitis remains 35-40%, both in resource-rich and resource-poor settings^{71,72}.

Currently used drugs. Only three classes of antifungal agents are currently used to treat cryptococcosis: polyenes (amphotericin B), azoles (fluconazole) and the pyrimidine analogue flucytosine (5FC) (**Figure 3**).

The cornerstone of treatment of cryptococcal meningoencephalitis is amphotericin B deoxycholate (AmBd), developed in the 1950s, which exerts its fungicidal effect both by binding to ergosterol in the cryptococcal cell wall (generating pores in the cell membrane) and by inducing cell death via oxidative damage⁷³⁻⁷⁵. AmBd is sometimes combined with 5-FC. The mechanism of action of 5-FC is deamination by the fungal enzyme cytosine deaminase into 5-fluorouracil (5-FU), which then acts via two pathways: 5-FU can be converted by cellular pyrimidine processing enzymes into 5-fluorodeoxyuridine monophosphate, which inhibits thymidylate synthetase and blocks DNA synthesis; or 5-FU can be converted into 5-fluorouridine triphosphate, which is incorporated into RNA, thereby disrupting protein synthesis and leading to growth arrest. AmBd and 5-FC act synergistically to produce the fastest rates of fungal clearance from CSF⁷⁶ and combination therapy results in a significant improvement in 10-week survival compared to treatment with AmBd alone⁷⁷. This combination remains the recommended 'gold standard' induction treatment in international treatment guidelines⁷⁸ but presents significant challenges in resource poor settings, since AmBd must be administered intravenously and has notable toxicities. In addition, neither AmBd nor 5-FC are widely available in countries where cryptococcosis is most prevalent⁷⁹.

To circumvent the problems associated with AmBd and 5-FC combination therapies, the combination of fluconazole with 5-FC (which can both be administered orally) and shorter (1-week) AmBd-based induction treatment is being compared to the standard 2-week induction regimens in a multi-site phase III African trial⁸⁰. Fluconazole is being tested because it has good oral bioavailability and excellent CSF penetration; these properties also make it recommendable for maintenance therapy after initial treatment. Fluconazole inhibits the fungal cytochrome P450 enzyme, 14 α -demethylase, which is required for conversion of lanosterol to ergosterol, an essential component of the fungal cell membrane. However, fluconazole is as a fungistatic (rather than

374 fungicidal) making it is less effective at pathogen clearance and not
375 recommended for initial therapy.

376
377 **Drug resistance.** Resistance to antimicrobials is a growing issue in infectious
378 disease and cryptococcosis is no exception. While environmental resistance is
379 rare, acquired resistance has been observed with all three classes of antifungals
380 in use against *Cryptococcus* species.

381 Polyene resistance is uncommon but has been reported in *C. neoformans*,
382 with mutations in sterol synthesis and therefore alteration of the target site noted
383 in isolates with extensive exposure to AmB ⁸¹. For 5-FC, single mutations at
384 varying points along the 5-FU intracellular pathways lead to *in vitro* and clinical
385 resistance. Therefore, monotherapy with 5-FC is not appropriate due to rapid
386 selection of resistant *Cryptococcus* leading to treatment failure; the drug is thus
387 always combined with either AmB or fluconazole. Fluconazole, like 5-FC, is
388 fungistatic, making it liable to evolution of secondary resistance during
389 prolonged treatment⁸². A key mechanism of resistance against fluconazole is the
390 selection of intrinsically resistant cryptococcal sub-populations ⁸³ that carry
391 specific chromosomal disomies ⁸⁴ and thus overexpress the *ERG11* gene (which
392 encodes the fluconazole target enzyme lanosterol-14 α -demethylase⁸⁵) or have
393 enhanced drug efflux by the ATP Binding Cassette (ABC) transporter-encoding
394 gene *C. neoformans* AntiFungal Resistance 1 (*CnAFR1*) ⁸⁶.

395
396 **New drugs.** Given the ongoing high global incidence and mortality from
397 cryptococcal meningoencephalitis, the dearth of drugs, together with toxicity and
398 the potential for development of resistance, there is an urgent need for new
399 drugs. Recent activity in this area has begun to highlight potential routes either
400 for the discovery of novel antifungals or for the repurposing of existing
401 molecules showing anti-cryptococcal activity (**Figure 3**).

402 An ideal antifungal drug should be fungal-specific, to avoid host cell
403 toxicity; this is challenging, given that fungal cellular processes are more closely
404 related to mammals than those that are targeted by common antimicrobials,
405 such as the ones used to target bacterial pathogens. Furthermore, an ideal
406 antifungal drug should target either a virulence factor or a fungal component

essential for fungal viability. Such a drug should be fungicidal when used alone or when combined with the widely available fluconazole, should have good oral bioavailability (allowing it to be readily administered even in resource-poor settings) and be able to enter cryptococcal niches within the host (such as phagocytes and the CNS).

One obvious target of such a drug is the cryptococcal cell wall. Unfortunately, the latest class of antifungals active against the cell wall, the β -1,3-D-glucan synthase inhibitors (echinocandins), have no significant anti-cryptococcal activity. However, synthesis of another cell wall component, glycosylphosphatidylinositol (GPI)-anchored mannoproteins, is inhibited by the orally-active experimental molecule E1210, which has *in vitro* activity against *Cryptococcus* and other medically-relevant fungi (such as *Candida* and *Scedosporium* species) and is currently in pre-clinical development⁸⁷.

Further along the development pipeline is VT-1129, an orally-available ergosterol synthesis inhibitor which shows good CNS penetration and is fungicidal in murine models of *Cryptococcus* infection. VT-1129 blocks the activity of CYP51, an essential enzyme in the pathway to produce ergosterol, and is currently entering human clinical trials ⁸⁸. Also in Phase I trials is the arlyamidine T-2307, which targets the fungal mitochondrial membrane⁸⁹. T-2307 is a fungicidal injectable compound that shows comparable efficacy to AmB in murine models of infection.

Given the lack of market forces driving pharmaceutical development for a neglected disease such as cryptococcal meningoencephalitis, an alternative, cheaper and more expedient strategy in drug development is the repurposing of drugs not originally developed for antifungal use. Recently developed high-throughput screening techniques have advanced the repurposing effort. One such powerful tool is chemical-genetic profiling, whereby large collections of cryptococcal knockout mutants, for which the function of a particular pathway is compromised, are screened against a library of small molecules⁹⁰, and the growth behavior of the screened strain (i.e. increased or decreased susceptibility) is then recorded. This technique was recently performed with 1448 knockout mutants of *C. neoformans* and demonstrated distinct differences in drug susceptibility between this species and the model organism

Saccharomyces cerevisiae which, until now, has been the standard choice for such screens⁹⁰. As proof of principle, this approach has identified a number of molecules that synergize strongly with fluconazole to inhibit ergosterol synthesis in *C. neoformans* and which are now being further investigated for potential clinical applicability. Moreover, this method has the additional advantage of providing information on the mechanism of action of lead compounds and can therefore identify both potential new drugs and potential new drug targets.

A more classical approach is to screen for compounds that trigger fungal lysis (detected by the release of adenylate kinase, a cytosolic enzyme, into the medium) or alter ATP content ⁹¹ (a particularly effective approach for identifying compounds that are antifungal under starvation conditions). This strategy has identified a collection of off-patent drugs⁹² with anti-cryptococcal activity that are additive or synergistic with fluconazole. These include drugs as diverse as amiodarone (a cardiac anti-arrhythmic drug), phenothiazines (widely used antipsychotics) and tamoxifen (an estrogen antagonist used in the treatment of breast cancer). Illustrating the utility of these approaches, tamoxifen in combination with fluconazole, decreased the *C. neoformans* burden in the brain by $\sim 1 \log_{10}$ CFU per gram of brain tissue, in a mouse model of infection⁹³. Finally, another candidate that has emerged from repurposing screens is the antidepressant sertraline (also known as Zoloft®), a drug that is fungicidal, has high CNS penetration, and appears to target fungal protein synthesis through an unknown mechanism⁹⁴. Sertraline is currently being evaluated in combination with AmBd and fluconazole in a phase II/III clinical trial ⁹⁵.

Outlook

The last five years have seen a remarkable revolution in our understanding of cryptococcosis. A deeper understanding of the natural ecology and an appreciation of the genetic and phenotypic diversity of this group of pathogens is transforming our understanding of cryptococcal pathogenesis. Meanwhile, huge progress has been made in understanding the host immune response to infection and how this process is hijacked by cryptococci to drive latency, dissemination and proliferation. However, despite these advances, cryptococcosis remains a

major worldwide killer, causing hundreds of thousands of deaths per year and the anti-cryptococcal drug arsenal remains limited. To address this, there is renewed focus on translational research to discover and develop new therapeutic agents and to evaluate new therapeutic strategies in a clinical setting. Whilst progress is being made in this respect, more is urgently required, and advances in understanding of the pathogenesis of *Cryptococcus spp* offer new opportunities for developing therapeutics beyond the traditional approaches of killing the fungal cell or preventing its replication. In particular, the rapidly expanding understanding of the *Cryptococcus*-host interface opens up new avenues for potential therapy development; for instance, in modifying host inflammatory responses, augmenting phagocytic clearance of the fungus, disrupting population signaling or preventing migration to the CNS. Together, such approaches offer the hope of significantly reducing the huge global burden of infection and making fatal cryptococcosis a disease of the past.

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Box 1. Clinical cryptococcosis.

Epidemiology. Since cryptococci are capable of extended latency within host cells⁴³ and most humans encounter the organism in early childhood⁸, it has been assumed that most clinical cases represent reactivation of a longstanding, asymptomatic infection (triggered, for instance, by falling CD4⁺ T-cell counts in HIV-infected individuals). The proportion of clinical disease representing reactivated latent disease versus primary infection is unknown in HIV-positive individuals, but a study in patients with cryptococcosis following solid-organ transplantation found only 52% of infections to be due to reactivation⁹⁶, suggesting that the classical view of cryptococcosis as a reactivating infection may not be accurate.

Emerging data are also highlighting the heterogeneity of cryptococcal disease worldwide, as illustrated by the prevalence of serum cryptococcal antigen (CrAg) in HIV-positive cohorts in different countries (see the figure, which displays the highest recorded prevalence per country). In addition, it is now clear that there is also considerable global heterogeneity in the fungal population structure. For example, *C. neoformans* var *grubii* (serotype A) is the predominant global cause of HIV-associated cryptococcal meningoencephalitis, but in China this organism frequently infects apparently immunocompetent hosts⁹⁷. Similarly, particular lineages of *C. neoformans* vary both in virulence⁹⁸⁻¹⁰⁰ and in their ability to infect immunocompromised or immunocompetent individuals¹⁰¹. In the near future, intensive whole genome sequencing efforts for both cryptococcal isolates and affected patients offers the possibility of being able to explain the relative contribution of host and pathogen genotypes underlying these global patterns of disease.

Susceptibility. In contrast to other systemic fungal infections (such as candidiasis), relatively little is known about genetic risk factors for cryptococcosis. However, recent allelic association studies have shown that apparently immunocompetent individuals with cryptococcosis are significantly more likely to have defects in mannose-binding lectin¹⁰² or be homozygous for the “232I” allele of the Fcγ receptor 2B (FcγR2B)¹⁰³, although these polymorphisms are relatively common and thus, on their own, are clearly not sufficient to render an individual fully susceptible to cryptococcosis. Therefore,

subtle defects in the innate immune response to fungi may underlie at least some cases of *C. neoformans* infection in otherwise healthy individuals. Similarly, in HIV-positive patients, allelic variation in a different FcγR, FcγR3A, also correlates with susceptibility¹⁰⁴. In this case, individuals with a higher affinity receptor variant are at greater risk of infection, perhaps indicating that efficient uptake of the pathogen may actually aid dissemination and drive more severe disease. This is particularly striking since the same is true from the pathogen perspective; cryptococcal strains that are more avidly phagocytosed drive more aggressive disease and carry a higher risk of death in patients¹⁰⁵. Thus, excessive phagocytosis as a result of either host or pathogen variation appears to drive cryptococcal dissemination, strongly supporting the “Trojan Horse” model of pathogen spread (see the main text).

Diagnosis. Diagnosis of cryptococcosis relies on detection either of the organism itself or its shed capsular GXM polysaccharide in serum or CSF. This has been hugely facilitated by the introduction of the point-of-care lateral flow cryptococcal antigen assay, which is cheaper and more sensitive than earlier serological tests¹⁰⁶. This test can detect very early dissemination and has facilitated cohort studies across the world, revealing a prevalence of cryptococcal antigens in HIV-infected patients ranging between 2 and 21%. As an increasing proportion of cases of cryptococcal meningoencephalitis are now presenting as “unmasking” of latent infection following therapy (i.e. the appearance of clinical symptoms following immune reconstitution by antiretroviral treatment), wider implementation of a ‘screen-and-treat’ approach is cost effective as a public health intervention and has been demonstrated to reduce mortality in African HIV cohorts in the first year on ART¹⁰⁷.

Box 2: The evolutionary history of cryptococci

The two *Cryptococcus* species, *C. gattii* and *C. neoformans*, probably diverged from a common environmental saprophyte ancestor around 30-40 million years ago^{108,109} (see the figure). For *C. neoformans*, extensive genetic data now indicates a common origin in sub-Saharan Africa^{5,110,111}. The observation that most non-African *C. neoformans* populations are near-clonal supports a model in which recombining African populations of cryptococci occasionally dispersed to

other parts of the globe. Coalescence analyses indicate that almost all of these events have occurred within the last 5000 years, suggesting the potential involvement of human or avian migrations in this process⁵.

Probing the origin and diversity of *C. gattii* has proven more challenging. There is a growing consensus that the evolutionary origins of this species lie within Australia and South America, since most dispersed lineages of *C. gattii* are near clonal (such as the lineage responsible for the Pacific Northwest Outbreak) but always cluster with Australian and South American isolates during phylogenetic analyses, with an estimated origin within the last 50 thousand years ¹¹²⁻¹¹⁴. A recurrent theme therefore appears to be that local populations of *C. gattii* in endemic areas (such as Brazil) undergo continual recombination, which occasionally results in a novel recombinant lineage that disperses and expands rapidly by means of clonal growth (either asexual cell division or same-sex mating) ^{112,113,115}.

Both species of *Cryptococcus* have a bipolar mating system in which cells are either mating type a (MATa) or mating type alpha (MAT α) (reviewed in ¹¹⁶). Classical mating involves genetic exchange between a MATa and MAT α strain, followed by normal Mendelian segregation of alleles. However, both species of cryptococci are also capable of same-sex mating in which two strains of the same mating type are able to exchange genetic material ^{115,117}. In addition, diploid and aneuploid strains are not uncommon^{118,119}, and inter- and intra-species hybrids can be found both in the environment and in patients^{120,121}. Thus the global population structure of these pathogens reflects a complex mix of “diversity generating” recombination and aneuploidy, coupled with highly clonal amplification steps during dispersion events.

Box 3: The evolution of virulence in cryptococci

Opportunistic pathogens represent an evolutionary enigma: why has natural selection driven the acquisition of often highly specific virulence factors when the majority of the population remain as exclusively environmental organisms for their entire existence? This conundrum is particularly pertinent for cryptococci, which are abundant in the environment and yet are remarkably well suited to survive in a human host.

A compelling hypothesis to resolve this conundrum is that of “accidental pathogenesis”¹²². This hypothesis proposes that cryptococcal pathogenesis does not result from direct selection for virulence within a mammalian host, but rather by the evolution of traits (which happen to be advantageous in mammals) in response to other selective pressures in both environmental and animal niches. So, for instance, the complex polysaccharide capsule, laccase activity and ability to synthesize melanin, which are all *Cryptococcus* virulence factors, are likely to offer protection against environmental pressures such as desiccation or exposure to ultraviolet light ¹²³, or aid in the colonization of plant hosts ¹²⁴. Similarly, cryptococci can replicate not only within vertebrate phagocytes, but also within free-living phagocytic amoebae¹²⁵ (see the figure). Despite the enormous evolutionary distance between vertebrates and amoebae, many of the mechanisms used by phagocytic white blood cells to kill pathogens (e.g. the generation of reactive oxygen species or secretion of antimicrobial peptides) are identical to those used by amoebae to digest ingested prey. Thus, over millions of years, cryptococci have been selected to evolve strategies that facilitate fungal growth and persistence within amoebae that coincidentally also enable their survival within phagocytes. Such strategies include not only stress-tolerance approaches, such as resistance to reactive oxygen species¹²⁶, but also elaborate mechanisms to regulate expulsion from host cells^{46,47}.

In addition, *Cryptococcus* has a remarkable ability to perturb adaptive immunity, preventing complete fungal clearance and resulting in latent infections.^{19,127}. Perhaps the ability to remain latent without perturbing its host is the strongest evidence for host adaptation by *Cryptococcus*. Since only higher vertebrates have adaptive immune systems, *Cryptococcus* species probably evolved these properties under the selective pressures of reptilian, avian or mammalian hosts within the environment, which also explains the diverse range of animals that can succumb to cryptococcosis.

Taken together, these observations suggest that interactions with both soil microorganisms (such as amoebae and nematodes) and vertebrates likely have a critical role in the virulence potential of *Cryptococcus* (reviewed in ¹²⁸). Intriguingly, laboratory studies have shown that selection pressure by amoebae can rapidly select for resistant, pseudohyphal forms of cryptococci²³. These

forms are attenuated in mammalian hosts and consequently frequently revert to yeast upon entry into a vertebrate host. Thus, rapid microevolutionary events may have an important role in driving cryptococcal pathogenesis in different hosts.

The paradigm of ‘accidental pathogenesis’ extends beyond cryptococci to other fungal¹²⁹ and even bacterial pathogens¹³⁰, such as *Aspergillus*, *Blastomyces* and *Legionella* species, and highlights two important issues. Firstly, as pathogens adapt to changing environments due to global warming, we may see additional instances of “accidental pathogenesis” through the selection of new traits that promote both environmental survival and pathogenesis in humans. Secondly, we should be alert to the fact that changes in human behavior and habitat use (e.g. increased tourist access to remote rainforest or desert areas) may expose us to novel potential pathogens that have been predisposed to infection via selection through environmental predators.

Box 4. Host immunity: too little or too much?

Poor inflammatory responses to cryptococci, such as those in patients with advanced HIV infection, lead to life-threatening meningoencephalitis. Consequently, immune profiling of patient peripheral T-cell responses and CSF cytokines has shown that those mounting a pro-inflammatory immune response are more likely to clear the pathogen and survive infection⁶⁶. Moreover, augmenting pro-inflammatory immune responses using adjunctive IFN- γ improves fungal clearance¹³¹. Conversely, individuals producing anti-cytokine antibodies that interfere with appropriate inflammatory responses are known to be at enhanced risk of infection¹³².

Although a potent immune response to *Cryptococcus* is clearly essential for fungal clearance, too strong a response can also be harmful. For instance, a low level of anti-inflammatory activity driven by both Th2 and regulatory T cells^{133,134} prevents complete immune paralysis. This is also the case for the classical antifungal cytokine IL-17, which is essential for resistance to cryptococcosis¹³⁵ but whose effects must also be regulated by IL-23 in order to prevent damage to the host due to excessive inflammation¹³⁶. Thus a “successful” immune response to cryptococcal infection appears to be a complex blend of

Th1, Th2 and Th17 responses, which must be counter-regulated to prevent either runaway fungal growth or damaging levels of inflammation.

This critical role for “restraining” inflammatory signaling is particularly highlighted by the problem of immune reconstitution inflammatory syndrome (IRIS). This life-threatening inflammatory reaction occurs in some HIV-infected patients during antiretroviral therapy (ART) and is attributable to the newly reconstituted immune system “overreacting” to residual pathogen antigen. Consequently, the timing of clinical intervention is critical; early introduction of ART is important to restore cell-mediated immunity, but if introduced too early (during the initial 2 weeks following induction of antifungal treatment) at a time of high fungal load, the risk of death is increased¹³⁷. Development of IRIS is particularly likely in patients whose initial pro-inflammatory response to cryptococcal infection is poor, resulting in high residual antigen burden¹³⁸. Coupled with an exaggerated baseline CNS chemokine response, this results in aberrant CNS immune responses following ART initiation, resulting in IRIS.

Excessive inflammation can also occur following withdrawal of immune suppression in solid organ transplant recipients, as well as in apparently immunocompetent patients. In such situations, steroids are often administered alongside antifungals. It remains unclear, however, whether steroids are beneficial in other contexts: a multi-centre clinical trial to address this issue (investigating the effect of adjunctive dexamethasone in patients with HIV-associated cryptococcal meningoencephalitis) has been terminated early and results are awaited¹³⁹.

Figure 1. Inflammatory signaling in response to cryptococcal infection.

Cryptococci inevitably shed microbial molecules that contain pathogen associated molecular patterns (PAMPS). Such fungal molecules are typically cell wall or capsular components such as chitin, β -glucan or glucuronoxylomannan (GXM), which are detected by immune sentinel cells, most notably dendritic cells (DCs). DC activation then summons T-cell help, inducing CD4⁺ T-cells to secrete cytokines that induce a T helper cell 1 (Th1) response (such as interleukin (IL)-12 and IL-23). Th1 cells produce pro-inflammatory cytokines (such as IFN- γ) that ultimately control fungal infection. However, some fungal PAMPs can influence DC activation, including modulating the levels of MHC-II or NF- κ B signaling. This leads to the generation of a Th2 response (mediated by the production of cytokines such as IL-4 and IL-13); this anti-inflammatory environment impacts the ability of macrophages to mediate fungal clearance.

Figure 2. Infection establishment and dissemination within the human host.

Cryptococcal cells typically enter the human host through the lung. Here they are recognized by patrolling phagocytes but can avoid uptake either by growing into very large “Titan” cells, or by relying on the antiphagocytic properties of the fungal capsule. If uptake occurs, however, cryptococci are able to survive and persist within phagocytes. For most strains, a failure in host immune function is then required to allow intracellular proliferation. However, the unusual Pacific Northwest Outbreak (PNO) strains of *C. gattii* can proliferate within immunocompetent host cells by exploiting a poorly-characterized “Division of Labour” mechanism: in response to reactive oxygen species generated by the phagocyte, some cryptococcal cells acquire an unusual morphology characterized by extensive tubularisation of their mitochondria, which increases survival of neighboring cells (via a mechanism that remains unclear). *Cryptococcus* proliferation within phagocytes ultimately leads either to host cell lysis or to a novel non-lytic escape mechanism termed vomocytosis. Upon replication in the lung, cryptococci are able to disseminate to other tissues, including the central nervous system (CNS). Entry into the CNS can occur in three ways: by squeezing between host endothelial cells (paracytosis), which

involves the fungal protease Mpr1; by moving directly through endothelial cells (transcytosis), in a process that is mediated by hyaluronic acid in the fungal capsule and the host receptor CD44; or by “hitching a ride” within migrating phagocytes, in a process termed the “Trojan horse” hypothesis.

Figure 3: Current and future therapies for cryptococcosis. Schematic representation of a cryptococcal cell, showing key current and potential therapeutic targets and examples of antifungal drugs acting at each site. Drugs in current clinical use are shown in red, novel drugs are shown in blue and repurposed drugs are shown in green. The three classes of antifungal agents currently used to treat cryptococcosis are polyenes (amphotericin B), azoles (fluconazole) and the pyrimidine analogue flucytosine (5-FC). Amphotericin B deoxycholate (AmBd) acts by binding to ergosterol in the cryptococcal cell wall, generating pores in the cell membrane, and by inducing cell death via oxidative damage. 5-FC is deaminated by the fungal enzyme cytosine deaminase into 5-fluorouracil (5-FU), which then inhibits thymidylate synthetase and blocks DNA synthesis or is converted into 5-fluorouridine triphosphate, which is incorporated into RNA and disrupts protein synthesis. Fluconazole inhibits the fungal cytochrome P450 enzyme, 14 α -demethylase, which is required for conversion of lanosterol to ergosterol, an essential component of the fungal cell membrane. E1210 inhibits the synthesis of the cell wall component glycosylphosphatidylinositol (GPI)-anchored mannoproteins. VT-1129 blocks the activity of CYP51, an essential enzyme in the pathway to produce ergosterol. The arylamidine T-2307 targets the fungal mitochondrial membrane. Tamoxifen (an estrogen antagonist used in the treatment of breast cancer) targets calmodulin and the anti-depressant sertraline appears to target fungal protein synthesis through an unknown mechanism.

753 **GLOSSARY**

754 *Pacific Northwest Outbreak* – an unusual cluster of cryptococcal disease in
755 otherwise healthy (rather than immunocompromised) individuals. First
756 identified on Vancouver Island, British Columbia, in 1999 (and hence originally
757 called the Vancouver Island Outbreak), both the causative organism and cases of
758 human and animal disease have now expanded into mainland Canada and the
759 northwestern USA, prompting a renaming of the outbreak.

760
761 *Iatrogenic* – caused by medical treatment. For instance, infections due to
762 contaminated surgical instruments.

763
764 *Zoonotic* – a disease transmitted from non-human animals to people

765
766 *Diploid* – having two homologous sets of chromosomes, one from each parent

767
768 *Aneuploid* – having an ‘unbalanced’ set of chromosomes; for instance, having only
769 a single copy of one chromosome in an otherwise diploid genome.

770
771 *Polyploid* – having multiple (more than two) sets of homologous chromosomes

772
773 *Founder* – the initial (small) group of individuals that seeds a new population.
774 For instance, the inoculum that starts an infection, or the first individuals to
775 arrive on a new island habitat.

776
777 *Quorum sensing* – the regulation of gene expression or behavior in response to
778 changes in the local population size.

779
780 *Paracrine* – a signal that acts close to where it is produced; for instance, on
781 neighbouring cells.

782
783 *Filamentation* – the growth of an organism by elongation without division.

784
785 *MHC Class II* – molecules that are expressed on the surface of professional
786 antigen presenting cells (such as macrophages and dendritic cells) and present
787 extracellular antigens to the immune system to coordinate an immune response

788
789 *Th1/Th2 response* – A broad characterization of the differentiation of CD4⁺
790 helper T cells (Th). Th1 responses are generally provoked by intracellular
791 pathogens and Th2 responses are typically involved in the elimination of
792 parasitic worms, harmful allergic responses, and dampening of Th1-mediated
793 inflammation. In the context of cryptococcal infection, Th1 responses are widely
794 thought to be protective and Th2 responses are detrimental.

795
796 *Melanization* – the production of the dark, insoluble pigment melanin, which
797 provides protection from high energy radiation and reactive oxygen molecules.

798
799 *Blood-brain barrier* – a specialized endothelial barrier that prevents the entry of
800 cells or large molecules into the central nervous system

801

802 *Paracytosis* – transitioning between tissues by moving between, rather than
803 through, adjacent cells.
804
805 *Transcytosis* - transitioning between tissues by moving directly through cells,
806 rather than between adjacent cells.
807
808 *Hyaluronic acid* – an abundant, high molecular weight polysaccharide that forms
809 part of the extracellular matrix, particularly in neural tissue.
810
811 *Cerebrospinal fluid (CSF)* – a clear fluid produced in the brain which bathes the
812 central nervous tissue and is slowly turned over.
813
814 *Fungistatic* – an antimicrobial agent that prevents fungal growth, but does not
815 kill the organism
816
817 *Fungicidal* – an antimicrobial agent that kills fungi, rather than simply preventing
818 growth
819
820 *Coalescence analysis* – an evolutionary analysis method in which genetic drift is
821 “played backwards” in order to calculate common ancestry of individuals within
822 a population and thereby estimate lineage branch points within an evolutionary
823 phylogenetic tree.
824
825 *Bipolar mating* – a system to control sexual reproduction that relies on a single
826 genetic locus at which individual organisms can carry one of two alleles,
827 effectively generating a species with two sexes.
828
829 *Regulatory T-cell* – a type of T-cell that functions to regulate the immune system,
830 typically by suppressing the function of proinflammatory effector T-cells.

831
832

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Author bios

Kirsten Nielsen received her PhD from North Carolina State University and then joined the medical mycology community while pursuing post-doctoral training at Duke University. Kirsten is currently an Associate Professor in the Department of Microbiology and Immunology at the University of Minnesota, where her research program focuses on factors influencing fungal pathogenesis both in animal models and during human diseases.

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cryptococcosis, host genetic susceptibility to cryptococcal infection and the evolution and mechanisms of fluconazole resistance in *Cryptococcus* sp.

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Online Summary

- Cryptococcosis is a widespread opportunistic fungal infection of humans and other animals.
- *Cryptococcus* species that infect humans likely evolved as “accidental pathogens” in response to environmental selective pressure.
- Recent genomic analyses have highlighted the evolutionary history of *Cryptococcus* species and narrowed down the geographical origin of an unusual, hypervirulent outbreak.
- Despite being accidental pathogens, cryptococci display a remarkable ability to manipulate the human immune response in order to facilitate disease establishment and spread.
- Detailed *in vivo* and *in vitro* characterization of *Cryptococcus* species has started to elucidate the details of multiple mechanisms of pathogenesis that likely have important roles in disease severity. These include changes in fungal morphology, interactions with host phagocytes and mechanisms that allow *Cryptococcus* to disseminate from the lung to the CNS.
- Renewed efforts to develop improved therapeutic approaches have highlighted potential new drugs and potential new uses for old drugs in the fight against cryptococcal disease.

ToC blurb

Recent studies have elucidated multiple virulence mechanisms used by *Cryptococcus* to infect, disseminate within and ultimately kill their human host. In this Review, May *et al.* describe these recent developments in understanding host-fungal interactions, discuss how they affect disease severity and debate current and future therapeutic interventions against cryptococcosis.

Subject categories

Biological sciences / Microbiology / Fungi / Fungal pathogenesis

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1. Kwon-Chung, J. K., Boekhout, T., Fell, J. W. & Diaz, M. Proposal to conserve the name *Cryptococcus gattii* against *C. hondurianus* and *C. basillisporus* (Basidiomycota, Hymenomycetes, Tremello-mycetidae). *Taxon* **51**, 804-806 (2002).
2. Hagen, F. et al. Recognition of seven species in the *Cryptococcus gattii*/*Cryptococcus neoformans* species complex. *Fungal Genet Biol* **78**, 16-48 (2015).
3. Springer, D. J. et al. *Cryptococcus gattii* VGIII isolates causing infections in HIV/AIDS patients in Southern California: identification of the local environmental source as arboreal. *PLoS Pathog* **10**, e1004285 (2014).
4. Chowdhary, A., Rhandhawa, H. S., Prakash, A. & Meis, J. F. Environmental prevalence of *Cryptococcus neoformans* and *Cryptococcus gattii* in India: an update. *Crit Rev Microbiol* **38**, 1-16 (2012).
5. Litvintseva, A. P. et al. Evidence that the Human Pathogenic Fungus *Cryptococcus neoformans* var. *grubii* May Have Evolved in Africa. *PLoS One* **6**, e19688 (2011).
6. Baddley, J. W. et al. Transmission of *Cryptococcus neoformans* by Organ Transplantation. *Clin Infect Dis* **52**, e94-8 (2011).

- 939 7. Lagrou, K. et al. Zoonotic transmission of *Cryptococcus neoformans* from a
940 magpie to an immunocompetent patient. *J Intern Med* **257**, 385-388 (2005).
- 941 8. Goldman, D. L. et al. Serologic evidence for *Cryptococcus neoformans*
942 infection in early childhood. *Pediatrics* **107**, E66 (2001).
- 943 9. Giles, S. S., Dagenais, T. R., Botts, M. R., Keller, N. P. & Hull, C. M.
944 Elucidating the pathogenesis of spores from the human fungal pathogen *Cryptococcus*
945 *neoformans*. *Infect Immun* **77**, 3491-3500 (2009).
- 946 10. Springer, D. J., Saini, D., Byrnes, E. J., Heitman, J. & Frothingham, R.
947 Development of an aerosol model of *Cryptococcus* reveals humidity as an important
948 factor affecting the viability of *Cryptococcus* during aerosolization. *PLoS One* **8**,
949 e69804 (2013).
- 950 11. Velagapudi, R., Hsueh, Y. P., Geunes-Boyer, S., Wright, J. R. & Heitman, J.
951 Spores as infectious propagules of *Cryptococcus neoformans*. *Infect Immun* **77**, 4345-
952 4355 (2009).
- 953 12. Zaragoza, O. et al. The capsule of the fungal pathogen *Cryptococcus*
954 *neoformans*. *Adv Appl Microbiol* **68**, 133-216 (2009).
- 955 13. McDonald, T., Wiesner, D. L. & Nielsen, K. *Cryptococcus*. *Curr Biol* **22**,
956 R554-5 (2012).
- 957 14. Zaragoza, O. & Nielsen, K. Titan cells in *Cryptococcus neoformans*: cells with
958 a giant impact. *Curr Opin Microbiol* **16**, 409-413 (2013).
- 959 15. Okagaki, L. H. et al. Cryptococcal cell morphology affects host cell
960 interactions and pathogenicity. *PLoS Pathog* **6**, e1000953 (2010).
- 961 16. Zaragoza, O. et al. Fungal cell gigantism during mammalian infection. *PLoS*
962 *Pathog* **6**, e1000945 (2010).
- 963 17. Wiesner, D. L. et al. Chitin Recognition via Chitotriosidase Promotes
964 Pathologic Type-2 Helper T Cell Responses to Cryptococcal Infection. *PLoS Pathog*
965 **11**, e1004701 (2015).
- 966 18. Gerstein, A. C. et al. Polyploid Titan Cells Produce Haploid and Aneuploid
967 Progeny To Promote Stress Adaptation. *MBio* **6**, (2015).
- 968 19. Alanio, A., Vernel-Pauillac, F., Sturny-Leclercq, A. & Dromer, F.
969 *Cryptococcus neoformans* Host Adaptation: Toward Biological Evidence of
970 Dormancy. *mBio* **6**, e02580-14 (2015).
- 971 20. Feldmesser, M., Kress, Y. & Casadevall, A. Dynamic changes in the
972 morphology of *Cryptococcus neoformans* during murine pulmonary infection.
973 *Microbiology* **147**, 2355-2365 (2001).
- 974 21. Neilson, J. B., Fromtling, R. A. & Bulmer, G. S. Pseudohyphal forms of
975 *Cryptococcus neoformans*: decreased survival in vivo. *Mycopathologia* **73**, 57-59
976 (1981).
- 977 22. Wang, L., Zhai, B. & Lin, X. The link between morphotype transition and
978 virulence in *Cryptococcus neoformans*. *PLoS Pathog* **8**, e1002765 (2012).
- 979 23. Magditch, D. A., Liu, T. B., Xue, C. & Idnurm, A. DNA mutations mediate
980 microevolution between host-adapted forms of the pathogenic fungus *Cryptococcus*
981 *neoformans*. *PLoS Pathog* **8**, e1002936 (2012).
- 982 24. Lin, J., Idnurm, A. & Lin, X. Morphology and its underlying genetic
983 regulation impact the interaction between *Cryptococcus neoformans* and its hosts.
984 *Med Mycol* (2015).
- 985 25. Bouklas, T. & Fries, B. C. Aging as an emergent factor that contributes to
986 phenotypic variation in *Cryptococcus neoformans*. *Fungal Genet Biol* **78**, 59-64
987 (2014).

988 26. Bouklas, T. et al. Old *Cryptococcus neoformans* cells contribute to virulence
989 in chronic cryptococcosis. *MBio* **4**, e00455-13 (2013).

990 27. Jain, N. et al. Isolation and characterization of senescent *C. neoformans* and its
991 implications for phenotypic switching and the pathogenesis of chronic cryptococcosis.
992 *Eukaryot Cell* **8**, 858-866 (2009).

993 28. Lee, H., Chang, Y. C., Nardone, G. & Kwon-Chung, K. J. TUP1 disruption in
994 *Cryptococcus neoformans* uncovers a peptide-mediated density-dependent growth
995 phenomenon that mimics quorum sensing. *Mol Microbiol* **64**, 591-601 (2007).

996 29. Albuquerque, P. et al. Quorum sensing-mediated, cell density-dependent
997 regulation of growth and virulence in *Cryptococcus neoformans*. *mBio* **5**, e00986-13
998 (2014).

999 30. Wang, L., Tian, X., Gyawali, R. & Lin, X. Fungal adhesion protein guides
1000 community behaviors and autoinduction in a paracrine manner. *Proc Natl Acad Sci U*
1001 *S A* **110**, 11571-11576 (2013).

1002 31. Albuquerque, P. C. et al. *Cryptococcus neoformans* glucuronoxylomannan
1003 fractions of different molecular masses are functionally distinct. *Future Microbiol* **9**,
1004 147-161 (2014).

1005 32. Idnurm, A. & Heitman, J. Light controls growth and development via a
1006 conserved pathway in the fungal kingdom. *PLoS Biol* **3**, e95 (2005).

1007 33. Schoffelen, T. et al. *Cryptococcus gattii* induces a cytokine pattern that is
1008 distinct from other cryptococcal species. *PLoS One* **8**, e55579 (2013).

1009 34. Piccioni, M. et al. A purified capsular polysaccharide markedly inhibits
1010 inflammatory response during endotoxic shock. *Infect Immun* **81**, 90-98 (2013).

1011 35. Angkasekwinai, P. et al. *Cryptococcus gattii* infection dampens Th1 and Th17
1012 responses by attenuating dendritic cell function and pulmonary chemokine expression
1013 in the immunocompetent hosts. *Infect Immun* **82**, 3880-3890 (2014).

1014 36. Qiu, Y. et al. Immune modulation mediated by cryptococcal laccase promotes
1015 pulmonary growth and brain dissemination of virulent *Cryptococcus neoformans* in
1016 mice. *PLoS One* **7**, e47853 (2012).

1017 37. Davis, M. J. et al. Macrophage M1/M2 polarization dynamically adapts to
1018 changes in cytokine microenvironments in *Cryptococcus neoformans* infection. *mBio*
1019 **4**, e00264-13 (2013).

1020 38. Voelz, K., Lammas, D. A. & May, R. C. Cytokine signaling regulates the
1021 outcome of intracellular macrophage parasitism by *Cryptococcus neoformans*. *Infect*
1022 *Immun* **77**, 3450-3457 (2009).

1023 39. Muller, U. et al. Abrogation of IL-4 receptor-alpha-dependent alternatively
1024 activated macrophages is sufficient to confer resistance against pulmonary
1025 cryptococcosis despite an ongoing T(h)2 response. *Int Immunol* **25**, 459-470 (2013).

1026 40. Hardison, S. E. et al. Protective immunity against pulmonary cryptococcosis is
1027 associated with STAT1-mediated classical macrophage activation. *J Immunol* **189**,
1028 4060-4068 (2012).

1029 41. Flaczyk, A. et al. IL-33 signaling regulates innate and adaptive immunity to
1030 *Cryptococcus neoformans*. *J Immunol* **191**, 2503-2513 (2013).

1031 42. Chen, G. H. et al. Inheritance of Immune Polarization Patterns is linked to
1032 Resistance versus Susceptibility to *Cryptococcus neoformans* in a Mouse Model.
1033 *Infect Immun* **76**, 2379-2391 (2008).

1034 43. Coelho, C., Bocca, A. L. & Casadevall, A. The intracellular life of
1035 *Cryptococcus neoformans*. *Annu Rev Pathol* **9**, 219-238 (2014).

1036 44. Alvarez, M. & Casadevall, A. Cell-to-cell spread and massive vacuole
1037 formation after *Cryptococcus neoformans* infection of murine macrophages. *BMC*
1038 *Immunol* **8**, 10.1186/1471-2172 (2007).

1039 45. Ma, H., Croudace, J. E., Lammas, D. A. & May, R. C. Direct cell-to-cell
1040 spread of a pathogenic yeast. *BMC Immunol* **8**, 15 (2007).

1041 46. Ma, H., Croudace, J. E., Lammas, D. A. & May, R. C. Expulsion of live
1042 pathogenic yeast by macrophages. *Curr Biol* **16**, 2156-2160 (2006).

1043 47. Alvarez, M. & Casadevall, A. Phagosome Extrusion and Host-Cell Survival
1044 after *Cryptococcus neoformans* Phagocytosis by Macrophages. *Curr Biol* **16**, 2161-
1045 2165 (2006).

1046 48. Nicola, A. M., Robertson, E. J., Albuquerque, P., Derengowski Lda, S. &
1047 Casadevall, A. Nonlytic exocytosis of *Cryptococcus neoformans* from macrophages
1048 occurs in vivo and is influenced by phagosomal pH. *mBio* **2**, e00167-11 (2011).

1049 49. Okagaki, L. H. & Nielsen, K. Titan cells confer protection from phagocytosis
1050 in *Cryptococcus neoformans* infections. *Eukaryot Cell* **11**, 820-826 (2012).

1051 50. Smith, L. M., Dixon, E. F. & May, R. C. The fungal pathogen *Cryptococcus*
1052 *neoformans* manipulates macrophage phagosome maturation. *Cell Microbiol* (2014).

1053 51. Davis, M. J. et al. *Cryptococcus neoformans*-Induced Macrophage Lysosome
1054 Damage Crucially Contributes to Fungal Virulence. *J Immunol* **194**, 2219-2231
1055 (2015).

1056 52. Johnston, S. A. & May, R. C. The human fungal pathogen *Cryptococcus*
1057 *neoformans* escapes macrophages by a phagosome emptying mechanism that is
1058 inhibited by Arp2/3 complex-mediated actin polymerisation. *PLoS Pathog* **6**,
1059 e1001041 (2010).

1060 53. Erb-Downward, J. R., Noggle, R. M., Williamson, P. R. & Huffnagle, G. B.
1061 The role of laccase in prostaglandin production by *Cryptococcus neoformans*. *Mol*
1062 *Microbiol* **68**, 1428-1437 (2008).

1063 54. Evans, R. J. et al. Cryptococcal Phospholipase B1 Is Required for Intracellular
1064 Proliferation and Control of Titan Cell Morphology during Macrophage Infection.
1065 *Infect Immun* **83**, 1296-1304 (2015).

1066 55. Vu, K. et al. Invasion of the central nervous system by *Cryptococcus*
1067 *neoformans* requires a secreted fungal metalloprotease. *mBio* **5**, e01101-14 (2014).

1068 56. Shi, M. et al. Real-time imaging of trapping and urease-dependent
1069 transmigration of *Cryptococcus neoformans* in mouse brain. *J Clin Invest* **120**, 1683-
1070 1693 (2010).

1071 57. Olszewski, M. A. et al. Urease expression by *Cryptococcus neoformans*
1072 promotes microvascular sequestration, thereby enhancing central nervous system
1073 invasion. *Am J Pathol* **164**, 1761-1771 (2004).

1074 58. Chang, Y. C. et al. Cryptococcal yeast cells invade the central nervous system
1075 via transcellular penetration of the blood-brain barrier. *Infect Immun* **72**, 4985-4995
1076 (2004).

1077 59. Jong, A. et al. Involvement of human CD44 during *Cryptococcus neoformans*
1078 infection of brain microvascular endothelial cells. *Cell Microbiol* **10**, 1313-1326
1079 (2008).

1080 60. Jong, A. et al. Invasion of *Cryptococcus neoformans* into human brain
1081 microvascular endothelial cells requires protein kinase C- α activation. *Cell*
1082 *Microbiol* **10**, 1854-1865 (2008).

1083 61. Liu, T. B. et al. Brain inositol is a novel stimulator for promoting
1084 *Cryptococcus* penetration of the blood-brain barrier. *PLoS Pathog* **9**, e1003247
1085 (2013).

1086 62. Kechichian, T. B., Shea, J. & Del Poeta, M. Depletion of alveolar
1087 macrophages decreases the dissemination of a glucosylceramide-deficient mutant of
1088 *Cryptococcus neoformans* in immunodeficient mice. *Infect Immun* **75**, 4792-4798
1089 (2007).

1090 63. Charlier, C. et al. Evidence of a role for monocytes in dissemination and brain
1091 invasion by *Cryptococcus neoformans*. *Infect Immun* **77**, 120-127 (2009).

1092 64. Chen, Y. et al. The *Cryptococcus neoformans* transcriptome at the site of
1093 human meningitis. *mBio* **5**, e01087-13 (2014).

1094 65. Robertson, E. J. et al. *Cryptococcus neoformans* ex vivo capsule size is
1095 associated with intracranial pressure and host immune response in HIV-associated
1096 cryptococcal meningitis. *J Infect Dis* **209**, 74-82 (2014).

1097 66. Jarvis, J. N. et al. Cerebrospinal Fluid Cytokine Profiles Predict Risk of Early
1098 Mortality and Immune Reconstitution Inflammatory Syndrome in HIV-Associated
1099 Cryptococcal Meningitis. *PLoS Pathog* **11**, e1004754 (2015).

1100 67. Datta, K. et al. Spread of *Cryptococcus gattii* into Pacific Northwest region of
1101 the United States. *Emerg Infect Dis* **15**, 1185-1191 (2009).

1102 68. Harris, J. R. et al. *Cryptococcus gattii* in the United States: clinical aspects of
1103 infection with an emerging pathogen. *Clin Infect Dis* **53**, 1188-1195 (2011).

1104 69. Ma, H. et al. The fatal fungal outbreak on Vancouver Island is characterized
1105 by enhanced intracellular parasitism driven by mitochondrial regulation. *Proc Natl*
1106 *Acad Sci U S A* **106**, 12980-12985 (2009).

1107 70. Voelz, K. et al. 'Division of labour' in response to host oxidative burst drives
1108 a fatal *Cryptococcus gattii* outbreak. *Nat Commun* **5**, 5194 (2014).

1109 71. Brizendine, K. D., Baddley, J. W. & Pappas, P. G. Predictors of mortality and
1110 differences in clinical features among patients with Cryptococcosis according to
1111 immune status. *PLoS One* **8**, e60431 (2013).

1112 72. Siddiqi, O. K. et al. Molecular diagnosis of central nervous system
1113 opportunistic infections in HIV-infected Zambian adults. *Clin Infect Dis* **58**, 1771-
1114 1777 (2014).

1115 73. Anderson, T. M. et al. Amphotericin forms an extramembranous and
1116 fungicidal sterol sponge. *Nat Chem Biol* **10**, 400-406 (2014).

1117 74. Belenky, P., Camacho, D. & Collins, J. J. Fungicidal drugs induce a common
1118 oxidative-damage cellular death pathway. *Cell Rep* **3**, 350-358 (2013).

1119 75. Gray, K. C. et al. Amphotericin primarily kills yeast by simply binding
1120 ergosterol. *Proc Natl Acad Sci U S A* **109**, 2234-2239 (2012).

1121 76. Brouwer, A. E. et al. Combination antifungal therapies for HIV-associated
1122 cryptococcal meningitis: a randomised trial. *Lancet* **363**, 1764-1767 (2004).

1123 77. Day, J. N. et al. Combination antifungal therapy for cryptococcal meningitis.
1124 *N Engl J Med* **368**, 1291-1302 (2013).

1125 78. Perfect, J. R. et al. Clinical practice guidelines for the management of
1126 cryptococcal disease: 2010 update by the infectious diseases society of america. *Clin*
1127 *Infect Dis* **50**, 291-322 (2010).

1128 79. Loyse, A. et al. Cryptococcal meningitis: improving access to essential
1129 antifungal medicines in resource-poor countries. *Lancet Infect Dis* **13**, 629-637
1130 (2013).

1131 80. www.controlled-trials.com/ISRCTN45035509.

1132 81. Kelly, S. L. et al. Resistance to amphotericin B associated with defective
1133 sterol delta 8-->7 isomerase in a *Cryptococcus neoformans* strain from an AIDS
1134 patient. *FEMS Microbiol Lett* **122**, 39-42 (1994).

1135 82. Bicanic, T., Harrison, T., Niepieklo, A., Dyakopu, N. & Meintjes, G.
1136 Symptomatic relapse of HIV-associated cryptococcal meningitis after initial
1137 fluconazole monotherapy: the role of fluconazole resistance and immune
1138 reconstitution. *Clin Infect Dis* **43**, 1069-1073 (2006).
1139 83. Sionov, E., Chang, Y. C., Garraffo, H. M. & Kwon-Chung, K. J.
1140 Heteroresistance to fluconazole in *Cryptococcus neoformans* is intrinsic and
1141 associated with virulence. *Antimicrob Agents Chemother* **53**, 2804-2815 (2009).
1142 84. Sionov, E., Lee, H., Chang, Y. C. & Kwon-Chung, K. J. *Cryptococcus*
1143 *neoformans* overcomes stress of azole drugs by formation of disomy in specific
1144 multiple chromosomes. *PLoS Pathog* **6**, e1000848 (2010).
1145 85. Sionov, E., Chang, Y. C. & Kwon-Chung, K. J. Azole heteroresistance in
1146 *Cryptococcus neoformans*: emergence of resistant clones with chromosomal disomy
1147 in the mouse brain during fluconazole treatment. *Antimicrob Agents Chemother* **57**,
1148 5127-5130 (2013).
1149 86. Posteraro, B. et al. Identification and characterization of a *Cryptococcus*
1150 *neoformans* ATP binding cassette (ABC) transporter-encoding gene, CnAFR1,
1151 involved in the resistance to fluconazole. *Mol Microbiol* **47**, 357-371 (2003).
1152 87. Miyazaki, M. et al. In vitro activity of E1210, a novel antifungal, against
1153 clinically important yeasts and molds. *Antimicrob Agents Chemother* **55**, 4652-4658
1154 (2011).
1155 88. <http://www.viamet.com/products/vt-1129>.
1156 89. Shibata, T. et al. T-2307 causes collapse of mitochondrial membrane potential
1157 in yeast. *Antimicrob Agents Chemother* **56**, 5892-5897 (2012).
1158 90. Brown, J. C. et al. Unraveling the biology of a fungal meningitis pathogen
1159 using chemical genetics. *Cell* **159**, 1168-1187 (2014).
1160 91. Dehdashti, S. J. et al. A high-throughput screening assay for assessing the
1161 viability of *Cryptococcus neoformans* under nutrient starvation conditions. *Anal*
1162 *Bioanal Chem* **405**, 6823-6829 (2013).
1163 92. Butts, A. et al. A repurposing approach identifies off-patent drugs with
1164 fungicidal cryptococcal activity, a common structural chemotype, and
1165 pharmacological properties relevant to the treatment of cryptococcosis. *Eukaryot Cell*
1166 **12**, 278-287 (2013).
1167 93. Butts, A. et al. Estrogen receptor antagonists are anti-cryptococcal agents that
1168 directly bind EF hand proteins and synergize with fluconazole in vivo. *mBio* **5**,
1169 e00765-13 (2014).
1170 94. Zhai, B., Wu, C., Wang, L., Sachs, M. S. & Lin, X. The antidepressant
1171 sertraline provides a promising therapeutic option for neurotropic cryptococcal
1172 infections. *Antimicrob Agents Chemother* **56**, 3758-3766 (2012).
1173 95. clinicaltrials.gov/ct2/show/NCT01802385.
1174 96. Saha, D. C. et al. Serologic evidence for reactivation of cryptococcosis in
1175 solid-organ transplant recipients. *Clin Vaccine Immunol* **14**, 1550-1554 (2007).
1176 97. Fang, W., Fa, Z. & Liao, W. Epidemiology of *Cryptococcus* and
1177 cryptococcosis in China. *Fungal Genet Biol* (2014).
1178 98. Beale, M. A. et al. Genotypic Diversity Is Associated with Clinical Outcome
1179 and Phenotype in Cryptococcal Meningitis across Southern Africa. *PLoS Negl Trop*
1180 *Dis* **9**, e0003847 (2015).
1181 99. Litvintseva, A. P. & Mitchell, T. G. Most Environmental Isolates of
1182 *Cryptococcus neoformans* var. *grubii* (Serotype A) are Not Lethal for Mice. *Infect*
1183 *Immun* **77**, 3188-3195 (2009).

100. Wiesner, D. L. et al. Cryptococcal genotype influences immunologic response and human clinical outcome after meningitis. *mBio* **3**, e00196-12 (2012).
101. Khayhan, K. et al. Geographically structured populations of *Cryptococcus neoformans* Variety *grubii* in Asia correlate with HIV status and show a clonal population structure. *PLoS One* **8**, e72222 (2013).
102. Ou, X. T. et al. Genotypes coding for mannose-binding lectin deficiency correlated with cryptococcal meningitis in HIV-uninfected Chinese patients. *J Infect Dis* **203**, 1686-1691 (2011).
103. Hu, X. P. et al. Association of Fcγ receptor IIB polymorphism with cryptococcal meningitis in HIV-uninfected Chinese patients. *PLoS One* **7**, e42439 (2012).
104. Rohatgi, S. et al. Fc gamma receptor 3A polymorphism and risk for HIV-associated cryptococcal disease. *mBio* **4**, e00573-13 (2013).
105. Sabiiti, W. et al. Efficient phagocytosis and laccase activity affect the outcome of HIV-associated cryptococcosis. *J Clin Invest* **124**, 2000-2008 (2014).
106. Jarvis, J. N. et al. Evaluation of a novel point-of-care cryptococcal antigen test on serum, plasma, and urine from patients with HIV-associated cryptococcal meningitis. *Clin Infect Dis* **53**, 1019-1023 (2011).
107. Mfinanga, S. et al. Cryptococcal meningitis screening and community-based early adherence support in people with advanced HIV infection starting antiretroviral therapy in Tanzania and Zambia: an open-label, randomised controlled trial. *Lancet* **385**, 60164-60167 (2015).
108. Findley, K. et al. Phylogeny and phenotypic characterization of pathogenic *Cryptococcus* species and closely related saprobic taxa in the Tremellales. *Eukaryot Cell* **8**, 353-361 (2009).
109. Xu, J., Vilgalys, R. & Mitchell, T. G. Multiple gene genealogies reveal recent dispersion and hybridization in the human pathogenic fungus *Cryptococcus neoformans*. *Mol Ecol* **9**, 1471-1481 (2000).
110. Litvintseva, A. P. & Mitchell, T. G. Population genetic analyses reveal the African origin and strain variation of *Cryptococcus neoformans* var. *grubii*. *PLoS Pathog* **8**, e1002495 (2012).
111. Litvintseva, A. P., Lin, X., Templeton, I., Heitman, J. & Mitchell, T. G. Many globally isolated AD hybrid strains of *Cryptococcus neoformans* originated in Africa. *PLoS Pathog* **3**, e114 (2007).
112. Billmyre, R. B. et al. Highly recombinant VGII *Cryptococcus gattii* population develops clonal outbreak clusters through both sexual macroevolution and asexual microevolution. *mBio* **5**, e01494-14 (2014).
113. Hagen, F. et al. Ancient Dispersal of the Human Fungal Pathogen *Cryptococcus gattii* from the Amazon Rainforest. *PLoS ONE* **8**, e71148 (2013).
114. Engelthaler, D. M. et al. *Cryptococcus gattii* in North American Pacific Northwest: whole-population genome analysis provides insights into species evolution and dispersal. *mBio* **5**, e01464-14 (2014).
115. Fraser, J. A. et al. Same-sex mating and the origin of the Vancouver Island *Cryptococcus gattii* outbreak. *Nature* **437**, 1360-1364 (2005).
116. Idnurm, A. et al. Deciphering the model pathogenic fungus *Cryptococcus neoformans*. *Nat Rev Microbiol* **3**, 753-764 (2005).
117. Lin, X., Hull, C. M. & Heitman, J. Sexual reproduction between partners of the same mating type in *Cryptococcus neoformans*. *Nature* **434**, 1017-1021 (2005).

1232 118. Lin, X. et al. Diploids in the *Cryptococcus neoformans* serotype A population
1233 homozygous for the alpha mating type originate via unisexual mating. *PLoS Pathog*
1234 **5**, e1000283 (2009).

1235 119. Ni, M. et al. Unisexual and heterosexual meiotic reproduction generate
1236 aneuploidy and phenotypic diversity de novo in the yeast *Cryptococcus neoformans*.
1237 *PLoS Biol* **11**, e1001653 (2013).

1238 120. Lin, X. et al. alpha AD alpha hybrids of *Cryptococcus neoformans*: evidence
1239 of same-sex mating in nature and hybrid fitness. *PLoS Genet* **3**, 1975-1990 (2007).

1240 121. Bovers, M. et al. Unique hybrids between the fungal pathogens *Cryptococcus*
1241 *neoformans* and *Cryptococcus gattii*. *FEMS Yeast Res* **6**, 599-607 (2006).

1242 122. Casadevall, A. Evolution of intracellular pathogens. *Annu Rev Microbiol* **62**,
1243 19-33 (2008).

1244 123. Wang, Y. & Casadevall, A. Decreased susceptibility of melanized
1245 *Cryptococcus neoformans* to UV light. *Appl Environ Microbiol* **60**, 3864-3866
1246 (1994).

1247 124. Warpeha, K. M., Park, Y. D. & Williamson, P. R. Susceptibility of intact
1248 germinating *Arabidopsis thaliana* to human fungal pathogens *Cryptococcus*
1249 *neoformans* and *C. gattii*. *Appl Environ Microbiol* **79**, 2979-2988 (2013).

1250 125. Steenbergen, J. N., Shuman, H. A. & Casadevall, A. *Cryptococcus*
1251 *neoformans* interactions with amoebae suggest an explanation for its virulence and
1252 intracellular pathogenic strategy in macrophages. *Proc Natl Acad Sci U S A* **98**,
1253 15245-15250 (2001).

1254 126. Zaragoza, O. et al. Capsule enlargement in *Cryptococcus neoformans* confers
1255 resistance to oxidative stress suggesting a mechanism for intracellular survival. *Cell*
1256 *Microbiol* (2008).

1257 127. Garcia-Hermoso, D., Janbon, G. & Dromer, F. Epidemiological evidence for
1258 dormant *Cryptococcus neoformans* infection. *J Clin Microbiol* **37**, 3204-3209 (1999).

1259 128. Kronstad, J. W. et al. Expanding fungal pathogenesis: *Cryptococcus* breaks
1260 out of the opportunistic box. *Nat Rev Microbiol* **9**, 193-203 (2011).

1261 129. Steenbergen, J. N., Nosanchuk, J. D., Malliaris, S. D. & Casadevall, A.
1262 Interaction of *Blastomyces dermatitidis*, *Sporothrix schenckii*, and *Histoplasma*
1263 *capsulatum* with *Acanthamoeba castellanii*. *Infect Immun* **72**, 3478-3488 (2004).

1264 130. Bliska, J. B. & Casadevall, A. Intracellular pathogenic bacteria and fungi--a
1265 case of convergent evolution? *Nat Rev Microbiol* **7**, 165-171 (2009).

1266 131. Jarvis, J. N. et al. Adjunctive interferon- γ immunotherapy for the treatment
1267 of HIV-associated cryptococcal meningitis: a randomized controlled trial. *AIDS* **26**,
1268 1105-1113 (2012).

1269 132. Saijo, T. et al. Anti-granulocyte-macrophage colony-stimulating factor
1270 autoantibodies are a risk factor for central nervous system infection by *Cryptococcus*
1271 *gattii* in otherwise immunocompetent patients. *mBio* **5**, e00912-14 (2014).

1272 133. Grahner, A. et al. IL-4 receptor-alpha-dependent control of *Cryptococcus*
1273 *neoformans* in the early phase of pulmonary infection. *PLoS One* **9**, e87341 (2014).

1274 134. Schulze, B. et al. CD4(+) FoxP3(+) regulatory T cells suppress fatal T helper
1275 2 cell immunity during pulmonary fungal infection. *Eur J Immunol* **44**, 3596-3604
1276 (2014).

1277 135. Murdock, B. J., Huffnagle, G. B., Olszewski, M. A. & Osterholzer, J. J.
1278 Interleukin-17A enhances host defense against cryptococcal lung infection through
1279 effects mediated by leukocyte recruitment, activation, and gamma interferon
1280 production. *Infect Immun* **82**, 937-948 (2014).

- 1281 136. Szymczak, W. A., Sellers, R. S. & Pirofski, L. A. IL-23 dampens the allergic
1282 response to *Cryptococcus neoformans* through IL-17-independent and -dependent
1283 mechanisms. *Am J Pathol* **180**, 1547-1559 (2012).
- 1284 137. Boulware, D. R. et al. Timing of antiretroviral therapy after diagnosis of
1285 cryptococcal meningitis. *N Engl J Med* **370**, 2487-2498 (2014).
- 1286 138. Chang, C. C. et al. Cryptococcosis-IRIS is associated with lower
1287 cryptococcus-specific IFN-gamma responses before antiretroviral therapy but not
1288 higher T-cell responses during therapy. *J Infect Dis* **208**, 898-906 (2013).
- 1289 139. www.controlled-trials.com/ISRCTN59144167.
- 1290